

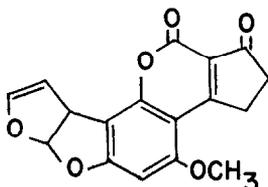
A NEW ROUTE TO
3a,8a-DIHYDROFURO[2,3-b]BENZOFURANS †

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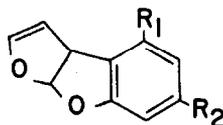
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The 3a,8a-dihydrofuro[2,3-b]benzofuran function occurs in several mold toxins, namely, the aflatoxins, sterigmatocystin, and versicolorin A¹. The toxic and carcinogenic properties of aflatoxin B₁ (1) are markedly dependent on the integrity of this function². Buchi and co-workers³ have devised a proficient synthesis for a 3a,8a-dihydrofuro[2,3-b]benzofuran (2b) as an intermediate in their ingenious construction of aflatoxin B₁. Since the system 2a and substituted



1



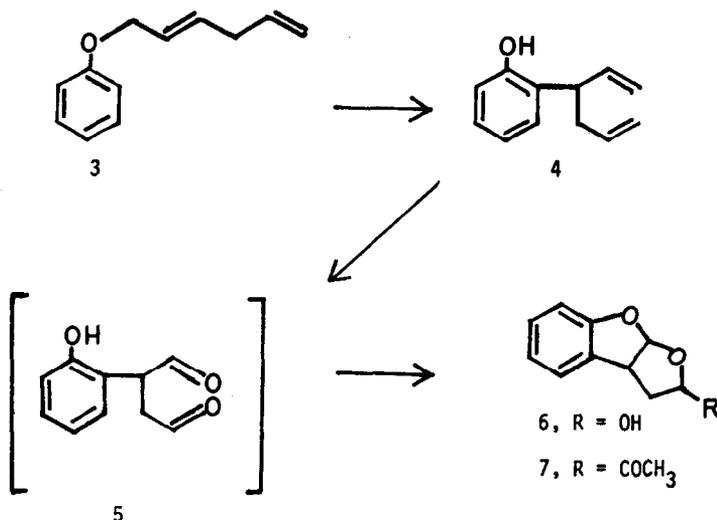
2a, R₁ = R₂ = H

2b, R₁ = OCOCH₃; R₂ = OCH₃

forms are desired for biological testing, we investigated an alternate route to 2a.

Alkylation of phenol in glyme with 1-bromo-2,5-hexadiene⁴ in the presence of potassium carbonate gave the primary ether 3. Fortunately, 3-bromo-1,5-hexadiene with phenol under the identical conditions yields the same ether, avoiding the necessity of separating the secondary bromide which accompanies its allylic isomers. When a mixture containing 19% of the secondary

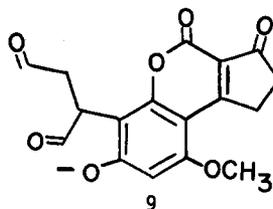
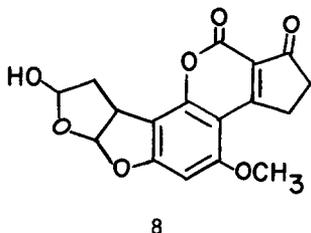
bromide is used, the ether produced contains less than one percent of its secondary isomer. The isomeric secondary ether can be preferentially formed by heating the phenol with neat secondary bromide.



Borontrichloride catalyzed Claisen rearrangement⁵ of the cold ether **3** leads to **4**, *o*-(3-hexa-1,5-dienyl)phenol, in 49% yield. The only side product from the rearrangement is the expected ether cleavage product, phenol. The traditional conditions for the Claisen rearrangement, heating (200° C) neat or in *N,N*-dimethylaniline, gave several unidentified products. Among these is probably a Cope rearranged product of **4** and the abnormal Claisen rearrangement product⁶.

Oxidative cleavage of the two double bonds in **4** with osmium tetroxide-sodium periodate yields the hemiacetal **6**, (ir(KBr pellet) 3470, 2940, 1601, 1489, 1257, 1222, 1190, 752, 744 cm⁻¹; λ max (EtOH) 278(2290), 273(2280), 115(6230); nmr (CDCl₃, δ) 7.16(4H, m), 6.45(1H, d, J=6H₂), 5.70 (1H, m), 4.15(1H, m), 2.50(2H, m; 1H, OH); mass spectrum 178(parent, 7%), 160(31), 147(30), 132(25) 131(100), 123(28), 103(29), 91(40), 89(27), 77(51), 51(31), 39(31). Hemiacetal **6** is easily converted to its more stable acetate **7**.

Concomitant formation of **6** from **5** is expected. 2-Allylphenol when cleaved by osmium tetroxide-sodium periodate spontaneously forms 2,3-dihydro-2-hydroxybenzofuran, which is easily converted to benzofuran by the action of phosphoric acid. Buchi^{3b} theorized that **9** was formed upon basic treatment of the hemiacetal of aflatoxin B₁, **8**, as evidenced by a bathochromic shift typical of a phenoxide in the ultraviolet spectra and by racemization of the natural material. Acidification of



9 regenerated optically inactive 8. Also treatment of 1 with aqueous acid, conditions which should establish a preferred equilibrium form among acetals and hemiacetal, produces 8^{3,7}.

Final confirmation of the structure of 6 and 7 is attained via synthesis by an alternate route, the procedure used by Buchi³ in his construction of 2b during his synthesis of Aflatoxin B₁, 1. Pyrolysis of 7 according to Buchi³ yields 2a.

Acknowledgment

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REFERENCES

- (1) J. V. Rodricks, J. Agr. Food Chem., 17, 457 (1969).
- (2) (a) G. N. Wogan, G. S. Edwards and P. M. Newberne, Cancer Res., 31, 1936 (1971). (b) J. L. Ayres, D. J. Lee, J. H. Wales and R. O. Sinnhuber, J. Nat. Cancer Inst., 46, 561 (1971).
- (3) (a) G. Buchi and Steven M. Weinreb, J. Am. Chem. Soc., 93, 746 (1971). (b) G. Buchi, et al., J. Am. Chem. Soc., 89, 6745 (1967).
- (4) J. C. H. Hwa and Homer Sims in "Organic Synthesis," Vol. 41, J. D. Roberts, Ed., John Wiley & Sons, New York, N.Y., 1961, pp. 49 and 51.
- (5) (a) W. Gerrard, M. F. Lappert and H. B. Silver, Proceedings Chem. Soc., 1957, 19. (b) P. Fahrni, A. Habich and H. Schmid, Helv. Chim. Acta, 43, 448 (1960).
- (6) E. N. Marvell, D. R. Anderson and Josephine Ong, J. Org. Chem., 27, 1109 (1962).
- (7) (a) A. E. Pohland, M. E. Cushmac and P. J. Andrellos, J. Ass. Offic. Anal. Chem., 51, 4 (1968)
(b) W. A. Pons et al., J. Am. Oil Chem. Soc., 49, 124 (1972).
- (8) D. S. Tarbell, K. J. H. Williams and E. J. Sehm, J. Am. Chem. Soc., 81, 3443 (1959).

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